Pharmacokinetic interactions between chloroquine, sulfadoxine and pyrimethamine and their bioequivalence in a generic fixed-dose combination in healthy volunteers in Uganda

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Abstract

Background: A pre-packaged fixed-dose formulation of chloroquine (CQ) and sulfadoxine/pyrimethamine (S/P) combination (Homapak) is widely used for the treatment of falciparum malaria in Ugandan children. It is however a product whose pharmacokinetics and interactions have not been studied.

Objectives: To explore possible pharmacokinetic interactions between CQ and S/P during co-administration, and to determine their bioavailability in the locally made Homapak compared to the Good Manufacturing Practice (GMP) made formulations.

Methods: Thirty-two adult healthy volunteers were randomized into four groups and given single oral doses of fixed-dose CQ+S/P combination (Homapak), or GMP formulations of S/P (Fansidar), CQ (Pharco), or their combination. Plasma samples were followed for 21 days, analysed by HPLC-UV methods, with pharmacokinetic modeling using the WinNonlin software.

Results: Sulfadoxine in Homapak was more rapidly absorbed (ka = 0.55 h^{-1}) than in

Fansidar + CQ (ka = 0.27 h^{-1} , p=0.004), but not more than S in Fansidar alone group (ka = 0.32 h^{-1} , p=0.03). No significant differences were observed in the other pharmacokinetic parameters of S, P and CQ when given together or separately. The relative bioavailability of CQ and S in Homapak showed bioequivalence to reference formulations.

Conclusions: There were no pharmacokinetic interactions between CQ, S and P when the compounds were given together, however, more investigations would be needed to explore this further. Compared with GMP made drugs, both S and CQ are bioequivalent in Homapak, the Ugandan made fixed-dose formulation. Furthermore, the absorption of S was more rapid which could be advantageous in malaria treatment.

Key words: sulfadoxine, pyrimethamine, chloroquine, bioequivalence, pharmacokinetic interactions *African Health Sciences* 2006; 6(2): 86-92

Introduction

In order to delay the development of resistance, combination therapy has been recommended for treatment of falciparum malaria¹. Several antimalarial drug combinations are now in use, but not all are affordable and easily available². Chloroquine (CQ) and sulfadoxine / pyrimethamine (S/P) are still the most affordable and easily available antimalarials, widely used in many resource poor nations despite the widespread resistance of *P. falciparum* to these drugs³.

The current antimalarial drug policy at health facilities in Uganda is Co-Artem, a co-formulated

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combination therapy containing artemether plus lumefantrine. However, the drug is not yet available⁴. A locally made pre-packaged fixed-dose generic formulation of chloroquine and sulfadoxine/pyrimethamine (locally known as Homapak)⁵ is still the recommended drug for the Home Based Management of Fevers (HBMF) program in the country⁶. The freely available Homapak has improved community access and compliance to prompt treatment of suspected malaria episodes in the under-five children, thereby reducing malaria related childhood mortality ^{7, 8}.

While generic formulations are promoted to address accessibility and affordability of drugs, reports on the quality of medicines in African countries have generally portrayed generics as poor quality or fake products⁹⁻¹². When the different products are either fake or substandard, the possibility of treatment failure and/ or interactions is potentially increased^{2, 13}. Co-administration of different products may also result in pharmacokinetic interactions¹⁴. Rengelshausen et al¹⁵ found that co-administration of methylene blue with chloroquine as treatment for malaria caused a 35% reduction in the area under the curve (AUC) of chloroquine. Grace et al¹⁶ have reported that chloroquine weakly inhibits the metabolism of arteether by inhibition of the enzyme arteether O-deethylase. Chloroquine has also been shown to decrease the bioavailability of ampicillin when they are co-administered¹⁷. The manufacturers of chloroquine advise that kaolin and antacids may reduce its absorption, and that cimetidine may inhibit chloroquine metabolism through the inhibition of CYP3A4, with the potential of increasing plasma CQ concentration. A Medline search did not yield any studies on sulfadoxine pyrimethamine or pharmacokinetic interactions.

The aim of this study was to explore possible pharmacokinetic interactions between CQ and S/P during co-administration in healthy adult Ugandan volunteers. Furthermore, we wanted to determine the bioequivalence of these compounds in the locally made Homapak compared to the Good Manufacturing Practice (GMP) made formulations.

Methods

Materials

The drugs used in the study, Fansidar®, Batch No. 3054 (Hoffman-La Roche, Basel, Switzerland) containing 500mg/25mg per tablet of sulfadoxine/pyrimethamine (SP) respectively, and Chloroquine Phosphate (CQ) Batch No. 174 (Pharco Pharmaceuticals, Alexandria, Egypt) equivalent to 150mg of the base, were bought from the National Medical Stores, Uganda. Both Fansidar and CQ (Pharco) are made according to Good Manufacturing Practice (GMP). Homapak[®] Batch No. 0402 (Kampala Pharmaceutical Industries Ltd. Kampala, Uganda) was provided by the Ministry of Health. Homapak is available as blister packs containing one tablet of S/P (500mg/25mg of sulfadoxine/pyrimethamine respectively), and three tablets of CQ (each containing chloroquine phosphate equivalent to 150mg of the base) fixed-dose.

Study design

The study was conducted at the Department of Pharmacology and Therapeutics, Faculty of Medicine, Makerere University, Kampala, Uganda. The flow chart (Figure 1) shows the recruitment, grouping, dosing and sampling of the volunteers. Only volunteers of proven good health, with no history of intake of antimalarials in the last three weeks, were included in the study. This was an open label study, in which the volunteers were randomly assigned to Homapak, Fansidar only, Fansidar+CQ or CQ only formulations. A two-stage randomization process was carried out, firstly to assign the medications into four groups, and secondly to assign the volunteers to a particular group, using computer generated random numbers.

In the morning of medication, each volunteer was given a single oral dose equivalent to sulfadoxine/ pyrimethamine =1500 mg/75 mgor chloroquine=600mg, singly or combined on an empty stomach, swallowed with a glass of drinking water under medical supervision. In order to obtain the above dosage for Homapak, three blister packs were opened, from which three tablets of S/P and four tablets of CQ were administered to each volunteer in the group (Fig. 1). Venous blood samples (2 x 5 ml) were collected from each volunteer in heparinized vacutainer tubes at intervals indicated in the chart (Figure 1). The samples were immediately centrifuged at 3500 g for 10 min to obtain plasma, which was then stored at -20°C until assay. Volunteers were allowed light breakfast (a cup of tea and a toast of bread) after the 2-hour sample. Thereafter, water was allowed *ad libitum*. Lunch was taken between 5 to 6 hours after drug administration.

HPLC Analysis

Drug plasma levels were assayed using HPLC at the Division of Clinical Pharmacology, Karolinska Institutet at Karolinska University Hospital, Huddinge, Stockholm, Sweden. Sulfadoxine (limit of determination 29μ mol/ l) and pyrimethamine (limit of determination 270nmol/ l) concentrations were determined using methods developed by Bergqvist et al¹⁸. Due to the huge differences in the plasma concentrations and the set limits of detection of sulfadoxine and pyrimethamine, separate runs with large volumes were used, leaving little plasma for pyrimethamine determination beyond the 24h period. Chloroquine concentrations (limit of determination 9.7nmol/l) were determined using the method described by Minzi et al¹⁹.

Statistical analysis and calculations

Pharmacokinetic calculations were performed by noncompartmental analysis using WinNonlin, version 4.1 (Pharsight Inc., Mountain View, CA, USA). Half-life ($t_{1/2}$) was calculated from the terminal slope ($\mathbf{\ddot{e}}$) of the concentration-time profile, and AUC was calculated using the trapezoidal rule, according to standard methods. Clearances (CL) and distribution volumes (V_d) were normalized by body weight. One-compartment model was used to calculate the absorption rate constant (k_a) of sulfadoxine and pyrimethamine, while a twocompartment model was applied for chloroquine. Relative bioavailability (AUC_{test}/AUC_{reference}) was determined from the ratio of the bioavailability of the drugs in Homapak (test) to that in the GMP drugs (reference). Bioequivalence was assumed when the relative bioavailability of the test drug was between 0.80-1.25 within the 90% confidence interval²⁰.

Data are presented as median values. All parameters were compared between treatment groups by the nonparametric Kruskal-Wallis test. To adjust for multiple comparisons, statistical significance was assumed only when p = 0.01. Statistical calculations were performed using Statistica version 6.1 (Statsoft Inc., OK, USA). The Student's t-test was used to compare the volunteer characteristics.

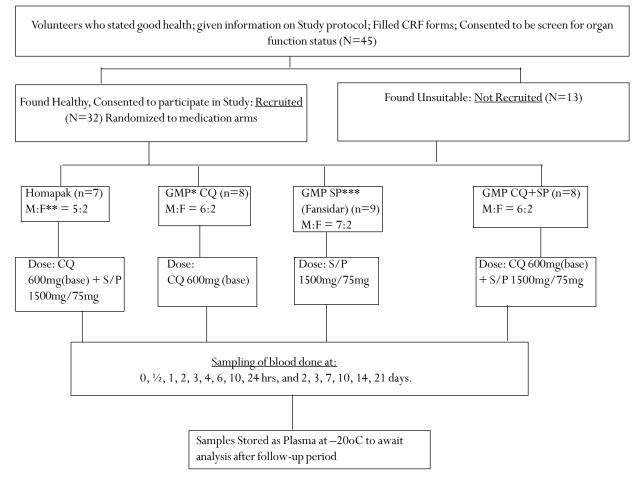
Ethical clearance

The Institutional Review Board of Makerere University, Uganda, and the Ethics Committee of Karolinska Institute, Sweden, approved the study (No.270/03). The Uganda National Council for Science and Technology (UNCST) gave permission to conduct the study. A signed, informed consent was obtained form each of the volunteers, and the study was performed in accordance with ICH-GCP guidelines as well as the Helsinki declaration.

Results

One volunteer in the Fansidar+CQ group had sulfadoxine concentration of 18.6 imol/l in plasma before drug administration and was therefore excluded. None of the remaining 31 volunteers had detectable amounts of S/P or CQ in the pre-dosing sample. Analysis of the characteristics of the volunteers showed there were no significant differences in age, body weight, liver function tests, and haemoglobin or creatinine levels between the groups. The drugs were generally well tolerated and no serious adverse reactions were reported. All volunteers could be sampled as scheduled on all occasions during the 21-day study period (Figure 1).

Figure 1: Study design showing recruitment, grouping, dosing and sampling of volunteers.



*Good Manufacturing Practices (GMP) formulations and combinations.

**M:F is the male:female ratio.

***One volunteer from this group was excluded from analysis when sulfadoxine was found in the pre-dosing plasma sample

Pharmacokinetics and bioavailability of sulfadoxine

Sulfadoxine in Homapak was more rapidly absorbed ($k_a = 0.55 h^{-1}$) than in Fansidar + CQ, ($k_{a=} 0.27 h^{-1}$, p=0.004) but not more than S in Fansidar alone group ($k_a = 0.32 h^{-1}$, p=0.03). The median absorption time (t_{max}) were 4h, 6h, and 10h for Homapak, Fansidar alone and Fansidar+CQ groups respectively. These observations were not significantly different between the groups.

Table 1: Pharmacokinetic parameters of sulfadoxine in the different formulations (median value and range).

	C _{max} (mol/l)	AUC ₀₋₂₁ (mmolh/l)	t _{max} (h)	t _{1/2} (h)	V _d (1/kg)	CL (ml/h/kg)	$\begin{array}{c} \mathbf{k}_{a} \\ (h^{-l}) \end{array}$	Relative Bioavailability ^a (90%CI)	AUC _{rest} % ^d
Formulation									
Fansidar	532	122	6	229	0.15	0.39	0.32	1.00	22
n=9	(455-649)	(102-159)	(1-6)	(136-272	2) (0.12-0.1	8) (0.30-0.56	5) (0.29-0.6	2) (Reference)	(7.6 – 26)
Fansidar+CQ	463	118	10	221	0.16	0.54	0.27	0.97	16
n=8	(332-546)	(99-140)	(6-24)	(154-347	7) 0.11-0.33) (0.45-0.58	3) (0.17-0.5	(0.88-1.06)	(11 – 32)
Homapak	530	131 ^b	4	224	0.15	0.43	0.55	1.07	21
n=7	(495-617)	(110-169)	(2-10)	(187-267	7) (0.14-0.20	0)(0.33-0.62)	0.42-2.6) ^c	(0.94-1.12)	(12 - 27)

^aThe acceptable value should be within 0.85 – 1.25 at 90% Confidence Interval

^b p=0.03 compared to Fansidar+CQ

 c p = 0.004, compared to Fansidar+CQ

 $^{d}AUC_{rest}$ is derived from AUC_{0.8} - AUC_{0.21} as a percentage of the observed AUC_{0.8}

No significant differences were observed in the other pharmacokinetic parameters (C_{max} , AUC, $t_{\frac{1}{2}}$, CL and V_{d}) for S in all the groups (Table 1 and Figures 2 and 3). With Fansidar as reference, the relative bioavailability of S in Homapak was 1.07 (90%CI: 0.94 – 1.12). The AUC_{rest} (AUC_{0.8} - AUC_{0.21}) for S was similar in both groups (21 % and 22% respectively). These results show that there is bioequivalence for S when given as Homapak compared with GMP made S/P (Fansidar).

Figure 2: Median plasma concentrations of sulfadoxine during 0-21 days after single dose of Homapak, Fansidar and Fansidar+CQ.

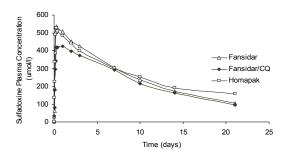
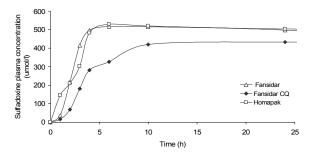


Figure 3: Median plasma concentrations of sulfadoxine during 0-24 hours after single dose of Homapak, Fansidar and Fansidar+CQ.

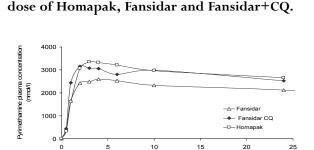


Pharmacokinetics of pyrimethamine

Pyrimethamine plasma concentrations could only be obtained during the absorption and early elimination phase. There were, however, no differences in the pharmacokinetic parameters of P (C_{max} , t_{max} and AUC_{0-24h}) in the first 24 hours between the groups (Table 2 and Figure 4).

Table 2: Pharmacokinetic parameters of pyrimethamine in the different formulations (median value and range).

Formulation	C _{max} ìmol/l	AUC _{24h} ìmolh/l	$t_{_{max}}$ h
Fansidar n=9	2.9 (2.0-3.9)	55 (43-69)	4 (2-6)
Homapak n=7	3.6 (2.6-4.8)	66 (54-80)	2 (2-4)
Fansidar+CQ n=8	3.3 (2.4-4.3)	63 (43-82)	4 (1-10)



Time (h)

Figure 4: Median plasma concentrations of

pyrimethamine during 0 - 24 hours after single

Pharmacokinetics and bioavailability of chloroquine

The median concentration-time profiles of chloroquine in the three groups were similar (Figure 5). No significant differences were found between the three groups for any of the pharmacokinetic parameters (Table 3). With GMP made CQ (Pharco) as reference, the relative bioavailability of CQ in Homapak was 0.97 (90%CI: 0.89 - 1.20). The AUC_{rest} for CQ in Homapak and CQ alone (Pharco) groups were similar (7.3 % and 6.8% respectively). These findings also show CQ in Homapak bioequivalent to GMP made CQ.

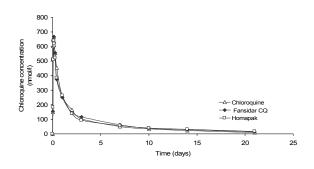
Table 3: Pharmacokinetic parameters of chloroquine in the different formulations (median values and range).

Formulation	C _{max} (mol/l)	AUC ₀₋₂₁ (mmolh/l)	t _{max} (h)	t _{1/2} (h)	V _d (1/kg)	CL (ml/h/kg)	$k_{a} (h^{-1})$	Relative Bioavailability ^a (90%CI)	AUC _{rest} % ^d
CQ ^a	760	34	2	155	113	0.50	1.8	1.00 (Reference	e) 7.3
n=8	(466-1186)	(19-54)	(1-4)	(87-232)	(55-257)	(0.39-0.77)	(0.27-3.4)		(1.9–21)
Fansidar+CQ	731	43	3	162	105	0.44	0.47	1.26	7.9
n=8	(449-1194)	(26-70)	(1-3)	(102-395)	(79-203)	(0.28-0.72)	(0.24-3.3)	(1.03 – 1.36)	(2.9 - 30)
Homapak	700	33	2	151	117	0.55	1.2	0.97	6.8
n=7	(495-813)	(17-45)	(1-4)	(93-325)	(69-548)	(0.38-1.2)	(0.19-7.8)	(0.89 – 1.20)	(1.7 – 24)

^aThe acceptable value should be within 0.85 - 1.25 at 90% Confidence Interval

 $^{\rm b}{\rm AUC}_{\rm rest}$ is derived from AUC_{0.8} - AUC_{0.21} as a percentage of the observed AUC_{0.8}

Figure 5: Median plasma concentrations of chloroquine during 0-21 days after single dose of Homapak, Chloroquine (GMP CQ) and Fansidar+CQ.



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Discussion

In the present study there were no clinically important pharmacokinetic interactions between chloroquine and sulfadoxine/pyrimethamine. This is an important finding, as the three compounds are used together as first line treatment of falciparum malaria in several resource poor nations^{3, 4}, even as more effective antimalarials are advocated^{21, 22}. A significant pharmacokinetic interaction between the compounds would have led to suppression or elevation of the bioavailability of any of the components, which was not the case in this study.

When administered as Homapak, S was more rapidly absorbed than as Fansidar, however, its bioavailability was found to be similar in the different formulations. This could probably have been due to different pharmaceutical preparations of the tablets^{13, 14}. It is well known¹³ that drugs in tablets forms with more rapid dissolution in the gastro-intestinal tract usually have more rapid systemic absorption, as measured by the absorption rate constant or time to maximal concentrations. Therefore, a rapid absorption, as was seen for S in Homapak, is desired for an antimalarial formulation for rapid parasite clearance, while slow absorption may lead to delayed parasite clearance and slower clinical response²³.

Impaired absorption may occur when different formulations are given simultaneously¹⁸. However, there was no difference in k_a when S was given as standard Fansidar with or without simultaneous intake of CQ. Although a reduction in absorption did not occur between Fansidar and CQ (Pharco) there is no guarantee that it cannot be seen when other preparations from other manufacturers are used, and as such more investigations need to be done in this area.

In poor resource countries locally made generic drugs are often used^{24, 25}. It is of importance to establish that they are not of substandard quality or even faked⁹⁻¹². Fortunately, in this study, Homapak has been found to be of good quality. Compared with GMP made formulations, both CQ and S in Homapak showed bioequivalence. Due to technical reasons, this could not be fully studied for P. There were, however, no differences in absorption of P during the first 24 hours. Based on these findings, we believe that there is probably good bioavailability of P in Homapak as well. However, in the absence of complete pharmacokinetic data on pyrimethamine, we cannot unequivocally conclude that Homapak is bioequivalent to GMP made drugs.

Conclusions

There were no pharmacokinetic interactions between CQ, S and P when the compounds were given together, however, more investigations would be needed to explore this further. Compared with GMP made drugs, both S and CQ are bioequivalent in Homapak, the Ugandan made fixed-dose formulation. Furthermore, the absorption of S was more rapid which could be advantageous in malaria treatment.

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Contribution by the different authors:

- Obua, C. (Corresponding author) was responsible for the conceptualization and design of the project, development of the project, execution of the experiments, data collection and analysis, and overall responsibility for the production of the manuscript.
- 2. Ntale M. was responsible for the pharmacokinetic analysis and scientific interpretation of results and input in the manuscript.
- 3. Lundblad, M.S. was responsible for the WinNonlin pharmacokinetic modeling, the scientific interpretation of the results, and input in the manuscript write-up.
- 4. Mahindi M. participated in the experimental design, laboratory analysis and gave critical comments on the manuscript.
- Gustafsson, L.L. Participated in the conceptualization and development of the project, and critical inputs during manuscript preparation.
- Ogwal-Okeng, J.W. participated in the conceptualization, laboratory and project development, design of the experiment and manuscript preparation.
- Anokbonggo, W.W. participated in experimental design, safety monitoring, critical revising of the manuscript.
- 8. Hellgren, U. together with the corresponding author, was responsible for the conceptualization and study design, overall ethical and scientific review of the project and manuscript preparation.

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